A Simplified Isolation Procedure of Four Cocarcinogenic Phorbolesters from Croton Oil

When earlier purification experiments succeeded in isolating 2 groups of compounds representing the total toxic, vesicant and cocarcinogenic potency of *Croton tiglium* L., they were classified groups A and B, respectively 1. Although the earlier separation procedure could be simplified in the meantime 2, so far no detailed information about the preparation of amounts sufficient for analytical and biochemical investigations was available. The following study precisely describes the isolation procedure leading to the pure cocarcinogenic compounds by 3 different steps only.

Methods. Test for cocarcinogenic action. 7-week-old³ albino mice (SaB) inbred for many generations at the Sandoz laboratories⁴ were used in all experiments. The manipulations were performed as described by Boutwell, Bosch and Rusch⁵ in a slightly modified manner ^{1,3,6}.

Isolation procedure. Preceding experiment: countercurrent distribution of 3.3 g croton oil DAB 67 in a Craig apparatus⁸ (49 tubes, upper phase and lower phase 10 ml each of the system¹ N-heptane-methanol-water = 1:1:0.1, 48 transfers) achieved separation into the fractions shown in Figure 1.

O'Keeffe countercurrent distribution: croton oil diluted with the upper phase (ratio 1:2) of the above heptane system was fed in 15-ml portions into tube 12 of an automatic 48-tube apparatus with a lower phase volume of 25 ml. About 95% of the croton oil remained in the upper phase, increasing its volume to 40 ml.

Column chromatography: an 8×150 cm column was filled with silicagel and eluted with an ethylacetate-chloroform-mixture (1:3). Up to 20 g of the resinous fraction can be applied to such a column.

Craig countercurrent distribution: the automatic apparatus with 200 tubes (both, lower and upper phase 25 ml) was fed once with up to 6 g of the active component A or B eluted from the column. The system carbontetrachloride-methanol-water = 2:1:0.3 was used and 1000 transfers performed.

Results. Cocarcinogenic activity. In all the mice pretreated with dimethylbenzanthracene, the compounds isolated stimulated during their 12 weeks application the appearance of papillomas up to 1.4 cm in diameter. The average values observed were 8.1 papilloma/mouse for A_1 , 6.5 for A_3 , 6 for B_1 , and 6.4 for B_2 .

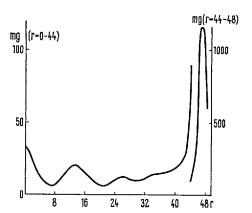
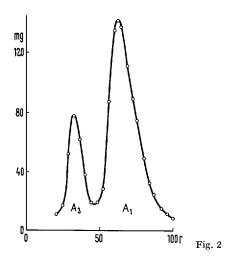


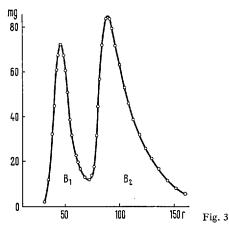
Fig. 1. Craig-countercurrent distribution (3.3 g croton oil, heptanesystem). Lower phase = upper phase = 10 ml, 48 transfers. Tubes 0-6 contain the resin, tubes 7-21 a colourless inactive oil, tubes 22-32 free fatty acids, tubes 33-48 neutral lipids. Abszissa: tube No. (fractions), ordinate: weights of the dried fractions in mg.

Isolation procedure. Preceding experiment: the fractions shown in Figure 1 contained the following mixtures: the active, resinous fraction with a partition coefficient p=0.05 was found in the tubes 0-6, representing 2.5% of the native croton oil; the tubes 7-21 consisted of a colourless, inactive oil, p=0.3, 3.5% of the oil; the tubes 22-32 contained the free fatty acids, p=1.2, 2%, and finally the neutral lipids were recovered in the tubes 33-48, p=15.4, 92%.

O'Keeffe distribution: dissolved in the upper phase, about 80% of the colourless oil, all the fatty acids and all

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Figs. 2 and 3. Craig-countercurrent distribution (carbontetra-chloride-system) of 4.4 g A (Figure 2) and 4.3 g B (Figure 3), respectively. Lower phase = upper phase = 25 ml, 1000 transfers. Abszissa: tube No. (fractions), ordinate: weights of the dried fractions in mg.

the lipids could be eliminated, i.e. about 97% of the inactive material of the original oil. Thin-layer chromatography (silicagel, ethylacetate-chloroform = 1:2) of the lower phase produced 9 distinct spots, representing the colourless oil (Rf = 0.9) and the active agents A (Rf = 0.25) and B (Rf = 0.35), mainly.

Chromatography: applying 12 g of the active resin obtained by evaporating the lower phase of the O'Keeffe distribution to the column, the active principles were found in fractions (100 ml each) 235–300 (B, 29% of the resin) and 345–405 (A, 16%). These components migrate as single spots in thin-layer chromatography 1.

Craig distribution: group A separated into the pure compounds A_1 (0.45% of the original croton oil) and A_3 (0.15%), group B into B_1 (0.4%) and B_2 (0.8%). The diagrams shown in Figures 2 and 3 are asymmetric, because the partition coefficients are dependent upon the concentration (Figure 4). This behaviour was most pronounced with compound B_2 . Consequently, the countercurrent distribution of B_2 resulted in the most asymmetric diagram. The effect decreases from B_2 - B_1 , from B_1 - A_1 ,

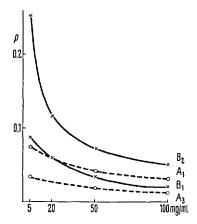


Fig. 4. Partition coefficients (ordinate) of the isolated compounds, determined in the carbontetrachloride-system. They show marked dependence on the concentration (abszissa) of the compounds.

and from A_1 – A_3 , respectively, as may be concluded from Figures 2 and 3 as well as from Figure 4.

Chemical characterization. IR-absorption spectra of the compounds isolated revealed hydroxylbonds, carbonyland esterbonds, and olefinic C=C absorptionbands. Hydrolysis liberated the following fatty acids: from A₁ $(C_{38}H_{56}O_8)$ myristic and acetic acid 10, from A₃ $(C_{38}H_{60}O_8)$ palmitic and acetic acid2, from B₁ (C₃₇H₅₈O₈) lauric acid and 2-methyl-butanoic acid and from B₂ (C₃₅H₅₄O₈) capric and 2-methyl-butanoic acid 11. All these compounds were shown to be diesters of the polyalcohol phorbol, C₂₀H₂₈O₆, analysed in 1934 by Flaschenträger¹². As phorbolesters are known to undergo alkaline hydrolysis easily 12,13 it may be of importance that the simplified procedure no longer uses the elimination of free fatty acids by 2Nalkalicarbonate, $pH = 10.95^{1,2,13,14}$. It is not yet experimentally excluded that this treatment may cause saponification and re-esterification 12, possibly thus leading to artifacts.

Zusammenfassung. Die schonende Reindarstellung von 4 kokarzinogenen Phorbolestern aus Krotonöl gelingt bereits nach 3 konsekutiven Trennoperationen (O'Keeffe-Gegenstromverteilung, Adsorptionschromatographie, Craig-Verteilung), deren Ausführung präzis beschrieben wird.

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